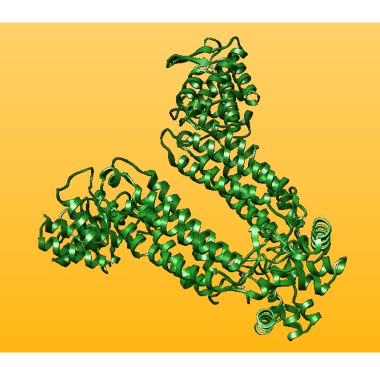
Oxygen Mediated Ketogenic Nanolubrication of ATP Synthase by Deuterium Depleted Low Viscosity Metabolic Water in Mitochondria

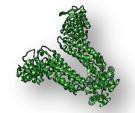
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Project	Special Issue "Medicinal Biochemistry of Deuterium Discrimination	ı" View project



Laszlo G. Boros, M.D.

Professor of Pediatrics (Endocrinology & Metabolism)

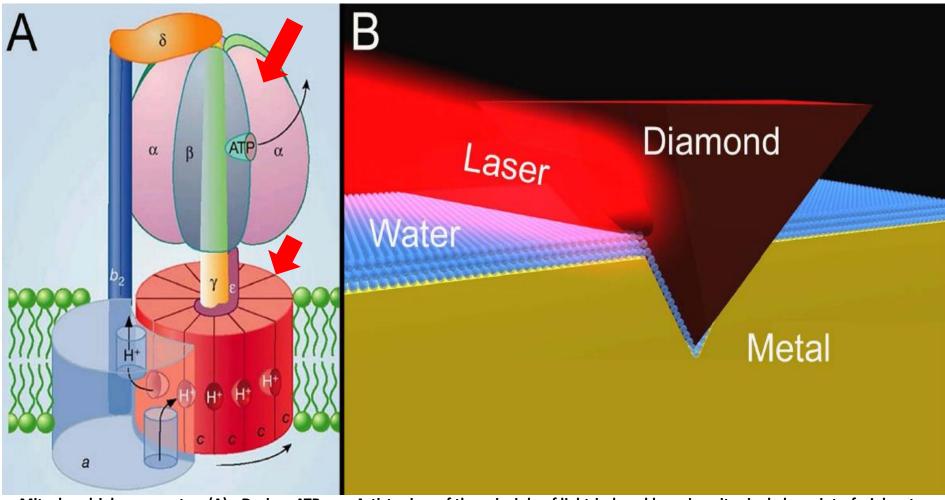
University of California Los Angeles
- UCLA School of Medicine,
Los Angeles, California, U.S.A.



OXYGEN MEDIATED KETOGENIC NANOLUBRICATION OF ATP SYNTHASE BY DEUTERIUM DEPLETED LOW VISCOSITY METABOLIC WATER IN MITOCHONDRIA

University of South Florida April 29, 2016

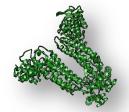
László G. Boros, M.D., Adjunct Professor of Pediatrics (Endocrinology & Metabolism), Harbor-UCLA Medical Center, Torrance, California, USA



Mitochondrial nanomotor (A): During ATP synthesis, the rotor turns about 9000 times per minute, where the lubricating water's viscosity controls velocity with significant deuterium induced kinetic isotope effects (Dorgan and Schuster 1981 J. Biol. Chem)

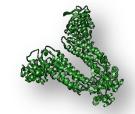
Artists view of the principle of light-induced low viscosity single layer interfacial water "flooding" a nanoindentation. Blue spheres are water molecules (H-O-H) forming the nanoscopic single water layer confined in the space between the diamond tip and nanoindentation from the direction of infrared light (B) (Sommer, et al. 2015 Nature Scientific Reports). Reprinted by permission from Macmillan Publishers Ltd: [NATURE] (Cross, 2004).

In this article: Deuterium depleted low viscosity single layer interfacial metabolic water "floods" natural nanoindentations (A; red arrows) of the ATP synthase protein as a natural lubricant, produced from low deuterium ketogenic substrates during complete oxidation, matrix water production, oxygen utilization and Complex-IV activity in mitochondria.



<u>www.CignatureHealth.com</u> ATP synthase movie







Laszlo G. Boros, MD

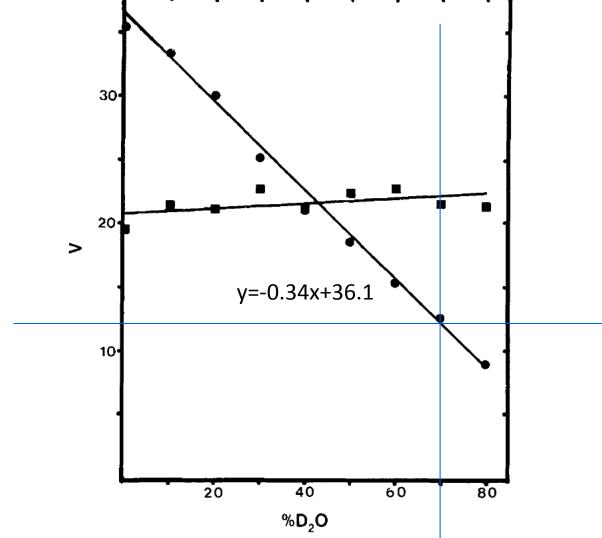


Fig. 2. Effect of D_2O on the rate (in micromoles/min/mg) of ATP (\blacksquare) and ITP (\blacksquare) hydrolyses by native F_1 . Nucleotide concentration is 5 mm.

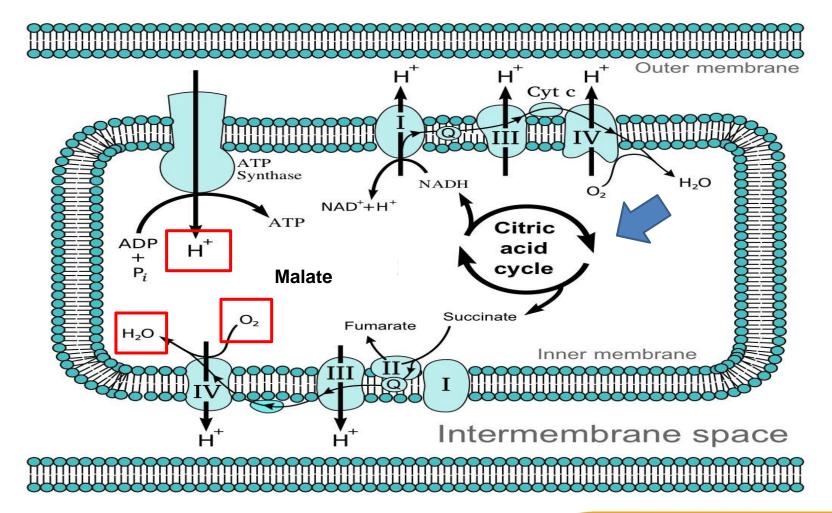
<u>Dorgan LJ</u>, <u>Schuster SM</u>. **The effect of nitration and D2O on the kinetics of beef heart mitochondrial adenosine triphosphatase.** <u>J Biol Chem</u>. 1981 Apr 25;256(8):3910-6.

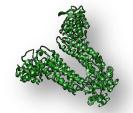
CYTOPLASMIC WATER IS WHAT WE DRINK AND SUPPLY THROUGH CIRCULATION



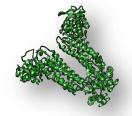
http://asiancorrespondent.com/92292/survey-what-worries-asias-kids/

MITOCHONDRIAL WATER IS WHAT IS PRODUCED IN THE MITOCHONDRIAL MATRIX

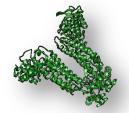




VCell TCA cycle Movie and popcorn (with butter and salt)



AMOUNTS AND PROPERTIES OF METABOLIC WATER PRODUCED IN MITOCHONDRIA DEPEND ON THE SUBSTRATES BEING OXIDIZED



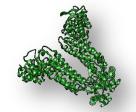
Nature 150, 21-21 (04 July 1942) | doi:10.1038/150021a0

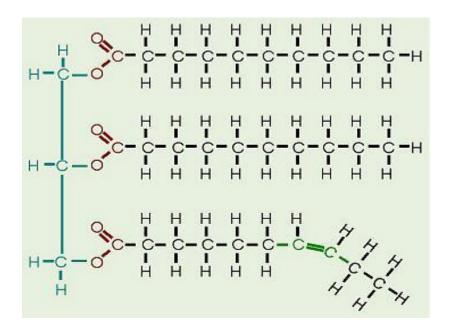
Metabolic Water and Desiccation

KENNETH MELLANBY

THE utilization by the body of ingested food substances and of tissue reserves yields among other things quantities of metabolic water. As the complete combustion of 100 gm. of fat produces about 110 gm. of metabolic water, whereas 100 gm. of carbohydrate yields only 55 gm. of water, fat reserves and fatty foods are believed to be particularly valuable as a protection against desiccation. This contention would appear to be supported by the fact that many animals which exist in deserts have large reserves of fat.

Laszlo G. Boros, MD

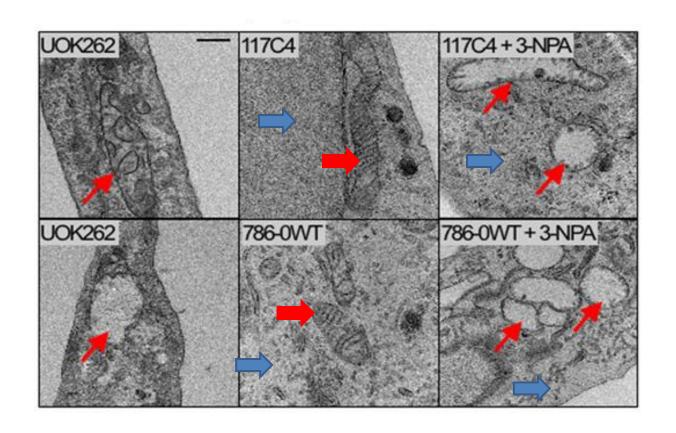


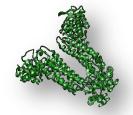


http://study.com/academy/lesson/what-is-fatty-acid-composition-structure-quiz.html

D-glucose

MITOCHONDRIAL AND CYTOPLASMIC WATER POOLS IN CANCER

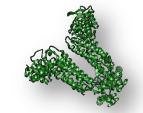




Let's produce water by oxidation of substrates using different food constituents

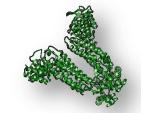
Let's turn the laboratory into "mitochondria"

This method will be transferred to USF



Methods I. (CuO catalysis)

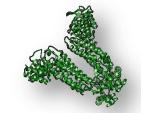
- Five mg samples are placed in Pyrex glass tubes and covered by 1 cm
 Copper(II) oxide (CuO) bead columns and dried under vacuum in batches of n=8 for at least 30 minutes (10⁻³ bar)
- Samples are completely oxidized in a 500 °C oven for 16 hours (Coleman-method) this is what happens in mitochondria and complex-IV
- The CuO slowly oxidizes all organic materials during the 16-hour period,
 followed by slow cooling down to room temperature
- One hundred mg zinc catalyst10 is measured in sealed Teflon tubes where water from the Pyrex tubes is carried by freezing at -80 °C this is what fumarate hydratase does -, while the CO2 product of complete substrate oxidation was removed by vacuum (cryogen separation) this is what we exhale through the lungs



Methods II. (IRMS)

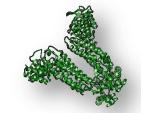
- Frozen water onto Zinc is reduced by Zn catalysis in 30 minutes in 500 °C ovens to H₂ gas this is what we transfer to NADP+
- ²H (D) ¹H ratio measurement using isotope ratio mass spectrometry (IRMS)
- Hydrogen gas using 2 microliter injections was loaded into the inlet of a Finnigan MAT delta S mass spectrometer at the Institute for Geological & Geochemical Research, Hungarian Academy of Sciences, Budapest, Hungary (Dr. István Fórizs)
- ²H₁ (D₁) concentrations (ppm) were calculated from the hydrogen's isotope composition

Harbor-UCLA Pediatrics
LABIOMED
HYD, LLC – MTA Geochemistry
SiDMAP, LLC



Methods III. (laser spectroscopy)

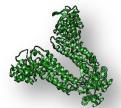
 Three laser spectroscope-validated BWS1, BWS2 and BWS3 laboratory standards, as well as one 29 ppm ²H-containing water sample (supplied by the vendor) were prepared the same way with the samples to control for recovery and guide calculations Harbor-UCLA Pediatrics
LABIOMED
HYD, LLC – MTA Geochemistry
SiDMAP, LLC



Methods IV. $(\delta^2 H)$

- Sample and standard isotopic compositions are expressed as $\delta^2 H$ values relative to V-SMOW according to the equation: $\delta = ((^2H/^1H sample ^2H/^1H standard)) / (^2H/^1H standard) x 1000 (‰), where <math>^2H/^1H$ ratios derive from either the sample or from the international standard of the European Atomic Energy Agency
- The precision of deuterium concentration ratios is less than 1.5 ppm based on duplicate (repeated) injections of samples

Deuterium content in eggs laid by 25 ppm DDW drinking hens



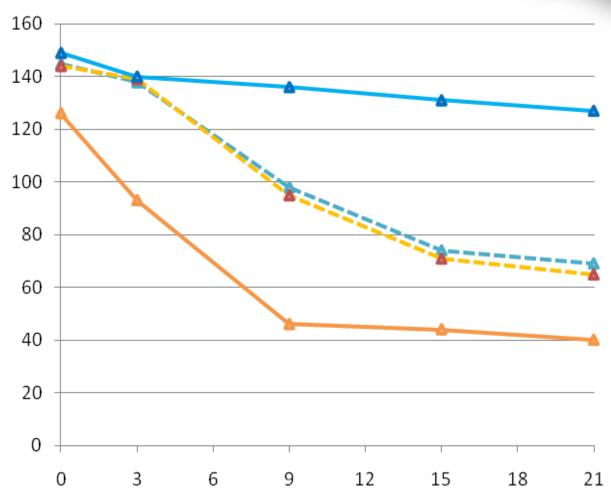
D content (ppm)

■ Water of egg white

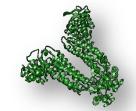
- △ - • Water of egg yolk

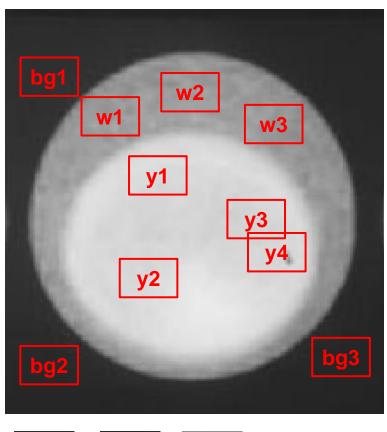
Organic bonds (white)

Organic bonds (yolk)



T1-WEIGHTED FRESH EGG





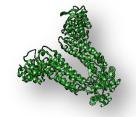
y1
y2
у3
y4
w1
w2
w3

		T1 fresh egg		
samples	variable	avg	min	max
white1	rgb	157	123	171
WillCi	hsb	62	48	67
white2	rgb	155	139	168
Willez	hsb	61	55	66
white3	rgb	153	108	171
William	hsb	60	42	67
yolk1	rgb	218	212	224
your	hsb	86	83	88
yolk2	rgb	217	211	229
yonce	hsb	85	83	90
yolk3	rgb	213	206	223
yomo	hsb	84	81	87
yolk4	rgb	213	137	226
your-	hsb	84	54	89
bg1	rgb	47	45	51
Dg.	hsb	18	18	20
bg2	rgb	41	38	49
29 <u>2</u>	hsb	16	15	19
bg3	rgb	38	36	40
bys	hsb	15	14	16

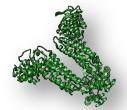
bg1 bg2 bg3

Size of samples: 60x40 pixel

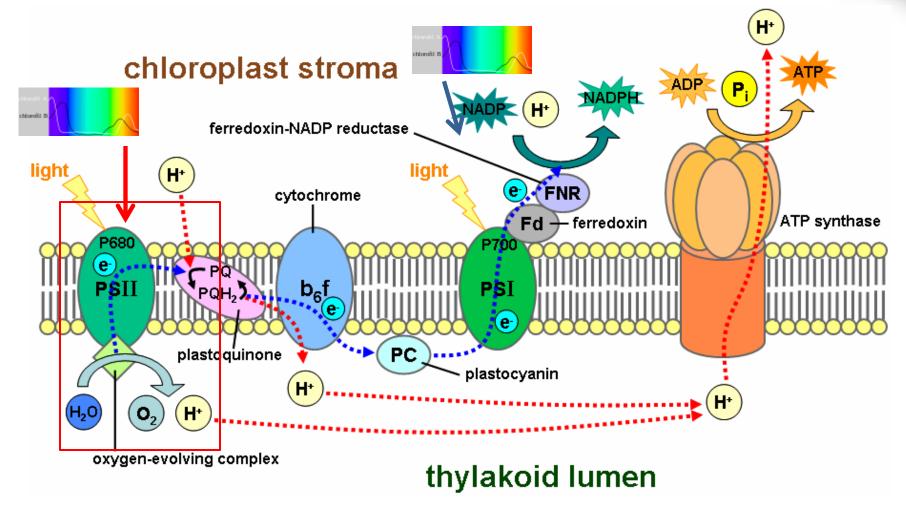
(1.59x1.06 cm)

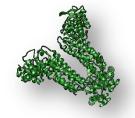


MITOCHONDRIAL WATER IS DEUTERIUM DEPLETED AND ITS LOW DEUTERIUM/HYDROGEN MOIETY ORIGINATES FROM ANIMAL FAT AND **GREEN PLANT SUGARS BY DEUTERIUM** DISCRIMINATION DURING **PHOTOSYNTHESIS**



Photosystem II - water-plastoquinone oxidoreductase





MITOCHONDRIAL WATER IS DEUTERIUM DEPLETED AND ITS LOW DEUTERIUM/HYDROGEN MOIETY SHOWS UP IN FAT/DNA BY REDUCTIVE SYNTHESIS



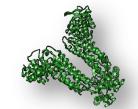


Can deuterium shield protons on MRI and alter longitudinal (thermal) proton relaxation properties





break 7x slower



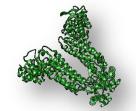
Single deuterium replacement disrupts collective proton tunneling of 96 other hydrogen molecules in hexagonal ice

Exceptional Isotopic-Substitution Effect: Breakdown of Collective Proton Tunneling in Hexagonal Ice due to Partial Deuteration

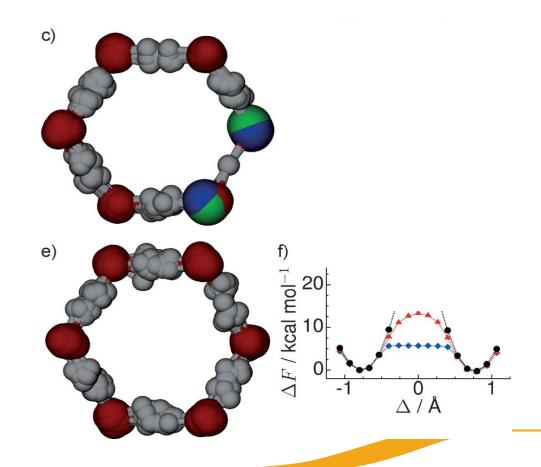
Christof Drechsel-Grau and Dominik Marx

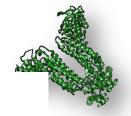
DOI: 10.1002/anie.201405989

Angewandte Chemie



Single deuterium replacement disrupts collective proton tunneling of 96 other hydrogen molecules in hexagonal ice





Interfacial water of cells and mitochondria

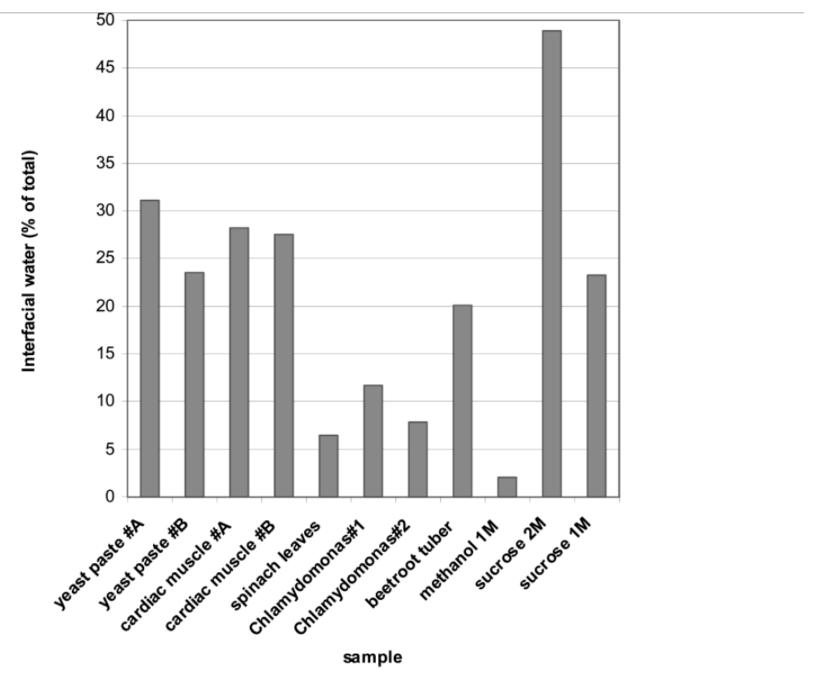


Inelastic Incoherent Neutron Scattering Measurements of Intact Cells and Tissues and Detection of Interfacial Water

Robert C. Ford,*,† Stuart V. Ruffle,‡ Anibal J. Ramirez-Cuesta,§ Ilias Michalarias,# Ilir Beta.# Aline Miller." and Jichen Li#

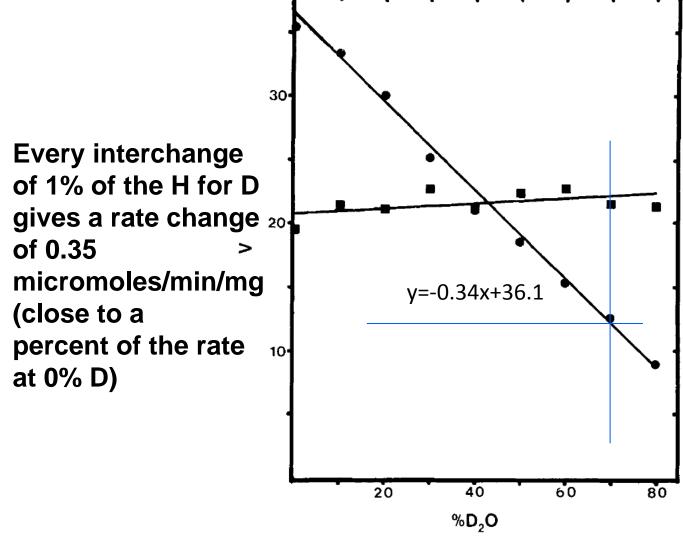
Contribution from the Departments of Biomolecular Sciences, Physics, and Chemical Engineering, UMIST, Manchester M60 1QD, U.K., School of Biological and Chemical Sciences, University of Exeter, Exeter EX4 4QG, U.K., and ISIS, Rutherford Appleton Laboratory, Chilton, Didcot, Oxfordshire, OX11 0QX, U.K.

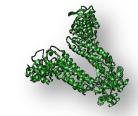
Received October 30, 2003; E-mail: r.ford@umist.ac.uk



Summary of the relative interfacial water content in the different cells and tissues surveyed, and a comparison to standard solutions of small interfacial water signal in the librational region of the IINS spectrum (45-130 meV) was integrated and then expressed as a percentage of the

In conclusion, we previously⁵ predicted that the interfacial water content in cells and tissues could represent a significant fraction of the total water, and in some cases (e.g., in mitochondria) could approach 100% of the water. We have now tested this prediction experimentally using IINS. An estimate of \sim 25–30% of water as interfacial was obtained from the data for yeast and cardiac cells. We expect this to be somewhat lower





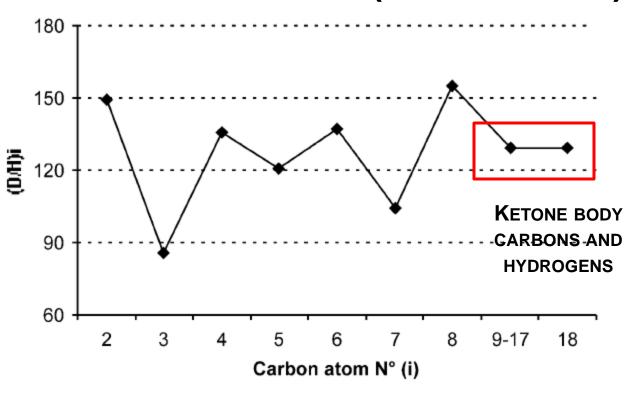
In deuterium free water the base ATP hydrolysis rate is calculated as 36.1 micromoles/min/mg in beef heart.

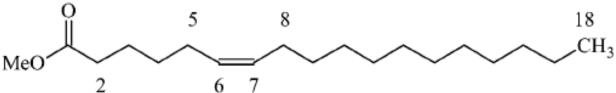
Fig. 2. Effect of D_2O on the rate (in micromoles/min/mg) of ATP (\blacksquare) and ITP (\blacksquare) hydrolyses by native F_1 . Nucleotide concentration is 5 mm.

<u>Dorgan LJ</u>, <u>Schuster SM</u>. The effect of nitration and D2O on the kinetics of beef heart mitochondrial adenosine triphosphatase. <u>J Biol Chem</u>. 1981 Apr 25;256(8):3910-6.

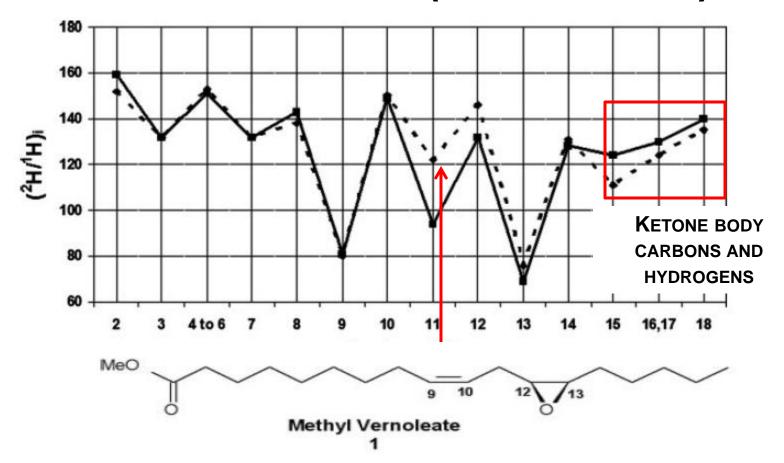
Source	Substrate, nutrient y=-0.34x+36.1 micromoles/min/mg	Average deuterium content [% decrease]*	Calculated change in ATP hydrolysis** with (%) increase to maximum rate***	Increase in ATP synthesis as the function of carbohydrate (flour) deriving metabolic water
Water (mean	International Atomic Energy	155.8		N/A
ocean water)	Agency Standard	[Reference range -		
		N/A]	+0	
Water of cottage	water from curd cheese -	151		
cheese (distilled	Quark dairy product	[-3.1]	+1.1** (3.04)***	0.8
by freezing				
under vacuum)				
Flour	wheat dehydrated all-	150		
	purpose PN-A-74022:2003	[-3.8]	+1.3 (3.6)	1 (Reference)
Table sugar	sucrose – Culinary white	146		
	refined (CAS 57-50-1)	[-6.3]	+2.1 (5.8)	1.6
Cottage cheese	curd - Quark dairy product	136		
(dry)		[-12.7]	+4.3 (11.9)	3.3
Sunflower oil	From Helianthus annus	130		
with ~84% high	seeds	[-16.6]	+5.6 (15.5)	4.3
oleic acid variety				
Butter	From cows' milk	124		
		[-20.4]	+6.9 (19.1)	5.3
Pork fat	Wet-rendered lard - pork	118		
	fatback	[-24.3]	+8.3 (22.9)	6.4

DOUBLE BONDS ARE ALMOST ALWAYS DEPLETED OF DEUTERIUM IN BIOLOGY (PARSLEY SEED)

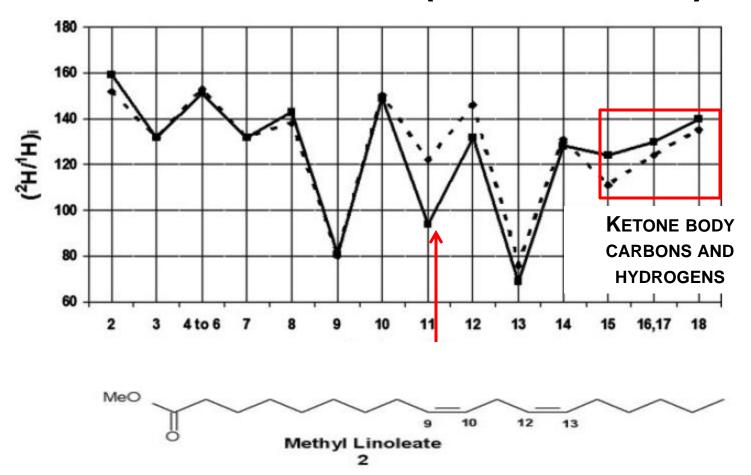




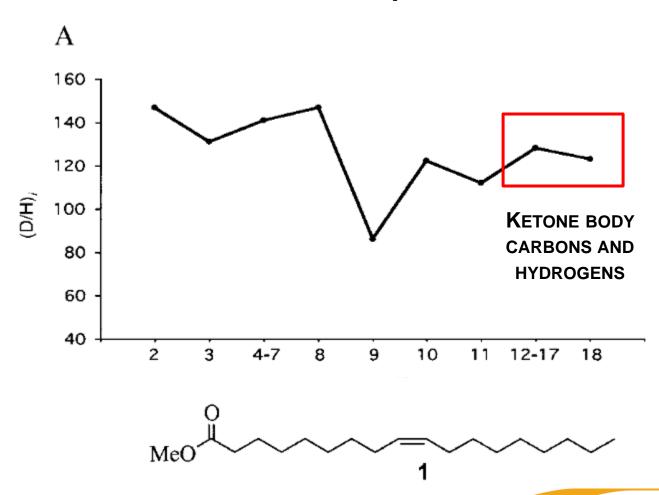
DOUBLE BONDS ARE ALMOST ALWAYS DEPLETED OF DEUTERIUM IN BIOLOGY (IRONWEED SEED)



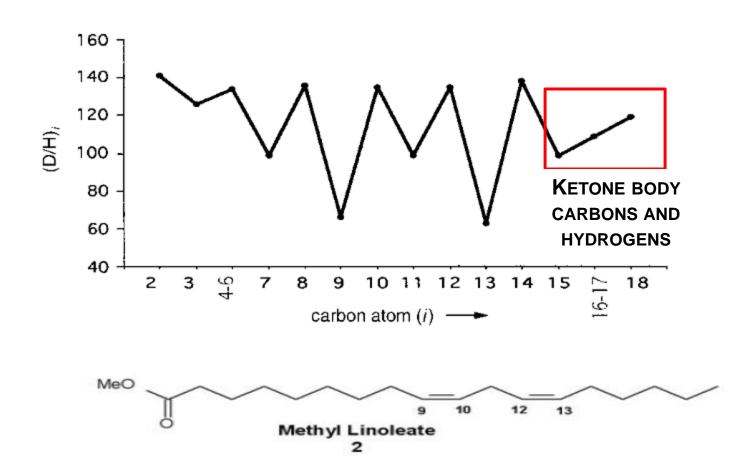
DOUBLE BONDS ARE ALMOST ALWAYS DEPLETED OF DEUTERIUM IN BIOLOGY (IRONWEED SEED)



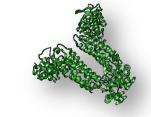
Double bonds are almost always depleted of deuterium in biology (Oleate - Sunflower)



Double bonds are almost always depleted of deuterium in biology (Sunflower)



Source (Species)	Fatty acids, carbon clusters and fragments y=-0.34x+36.1 micromoles/min/mg	Average deuterium content [% decrease]*	Calculated change in ATP hydrolysis** with (%) increase to maximum rate***	References
Ironweed (Vernonia galamensis)	vernoleate linoleate	127.5 [-18.2%] 123.9 [-20.5%]	+6.2 (17.1) +6.8 (18.8)	Billault, I., et al. 2005. J. Biol. Chem. 280, 17645–17651.
Peanut oil (Arachis hypogaea)	palmitate and stearate (cluster C1-C7)	119.9 [-23.1%]	+7.9 (21.9)	Duan, J.R., et al. 2002. ChemBioChem . 3, 752–759.
Sunflower (Helianthus annus)	linoleate (cluster C13-C18) terminal ketones	110 [-29.7%]	+10.1 (27.7)	Billault, I., et. al. 2001. ChemBioChem 2, 425–431.
Parsley seed (Petroselinu m crispum)	petroselinic acid (C18:1D ⁶) y=-0.34x	124.9 [-19.9%] +36.1	+6.8 (18.8)	Guiet, S., et al. 2003. Phytochemistry - 64, 227-33.



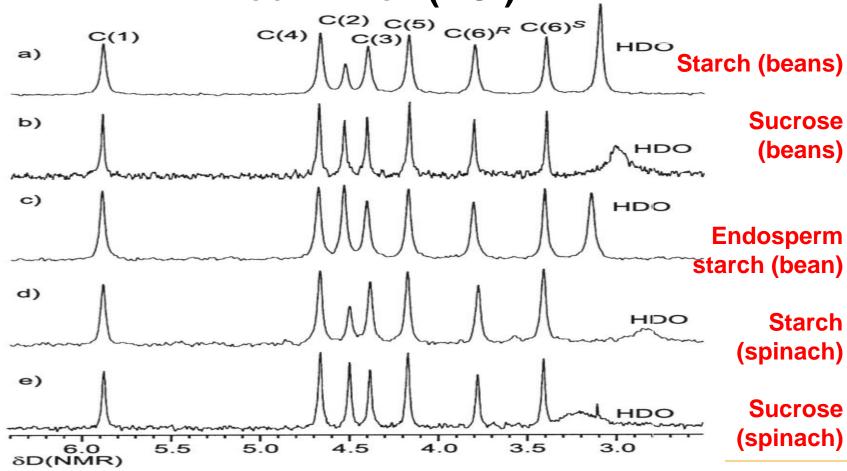
Plant, Cell & Environment Volume 22, Issue 5, pages 525–533, May 1999

Intramolecular deuterium distributions reveal disequilibrium of chloroplast phosphoglucose isomerase

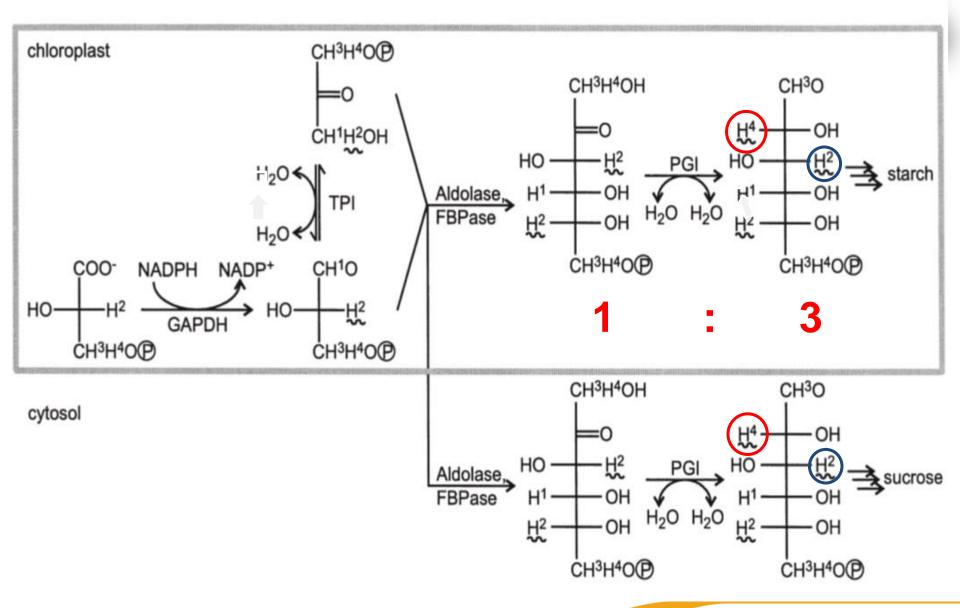
J. SCHLEUCHER, P. VANDERVEER, J. L. MARKLEY, T. D. SHARKEY

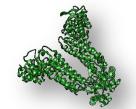
DOI: 10.1046/j.1365-3040.1999.00440.x

INTRAMOLECULAR DEUTERIUM DISTRIBUTIONS REVEAL DISEQUILIBRIUM OF CHLOROPLAST PHOSPHOGLUCOSE ISOMERASE (PGI)



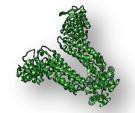






CLINICAL SIGNIFICANCE OF DEUTEROBOLOMICS DOI 10.1007/s11306-016-0961-5

- Submolecular oncology
- http://dx.doi.org/10.1016/j.mehy.2015.11.016



Acknowledgments

Dr. Howard Katz – Chair – Johns Hopkins Engineering

Dr. Andrei Sommer – *University of Ulm*

Dr. Gábor Somlyai – HYD, LLC

Dr. W-N Paul Lee – *UCLA*Agi Hirshberg – <u>www.Pancreatic.org</u>

Eszter Boros – H/D in tumors and visual art

Dr. Dominic D'Agostiono - USF Ron Penna – *Quest Nutrition and Epigenix* <u>www.epigenixfoundation.org</u>

